

Highly-degrading bacteriophages as a therapeutic treatment against *Pseudomonas aeruginosa* in cystic fibrosis

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BACKGROUND

- *Pseudomonas aeruginosa* can cause up to 20% of nosocomial infections [1].
- *P. aeruginosa* colonize the lung of cystic fibrosis patients.
- *Pseudomonas* infections are difficult to control because its resistance to many antibiotics and disinfectants, mainly due to efflux pumps and biofilm [2].
- Phage therapy has many advantages over antibiotics but needs further research.
- The use of genetically-modified phages is an interesting site-directed treatment.

INITIAL HYPOTHESIS AND OBJECTIVES

- A cocktail composed by *P. aeruginosa* bacteriophages overexpressing an alginate-degrading enzyme could be an effective treatment in cystic fibrosis patients colonized by this bacterium.
- The aim of this project is to design a phage cocktail based in several lytic and non-transducing phages encoding an alginate lyase, and evaluate its safety and effectiveness *in vitro* and *in vivo*.

EXPERIMENTAL DESIGN

1. The Step-by-step method (SBS) will be used for the selection of infective phages against each resistant strain that could appear. The method ends when last phage-resistant mutant is sensible to the first phage [3].

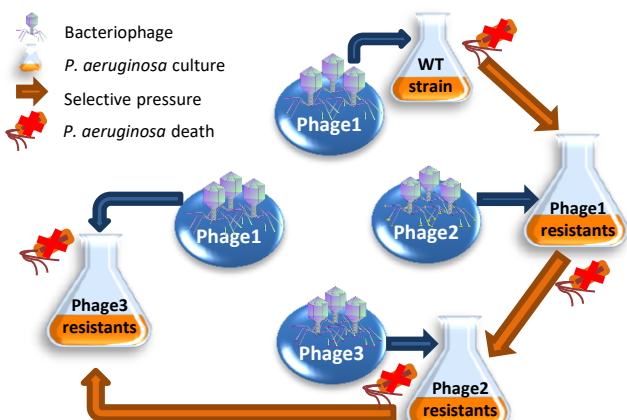


Figure1. Representation of the SBS method.

2. Bacteriophage Recombineering of Electroporated DNA (BRED) will be conducted for gene insertion in the phage genome. The genome and the desired gene are co-electroporated in a strain with a recombination system [4].

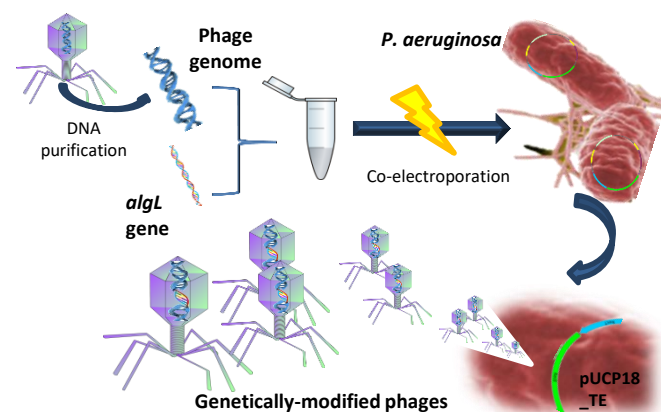


Figure2. Representation of BRED Technology.

3. *In vitro* biofilm and cell viability analysis will be performed in microtiter plates and continuous flow cells. Total biomass quantification and cell viability will be determined by Syto9 and FDA assays, respectively.

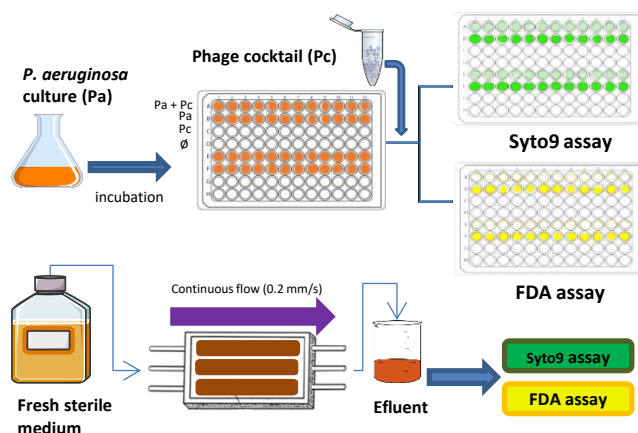


Figure3. Diagram of *in vitro* analysis of early (up) and mature (down) biofilms.

4. For *in vivo* determination of safety and effectiveness, survival rate of mice will be determined as well as quantification of cytokines, LDH, phages and bacteria in bronchoalveolar fluids.

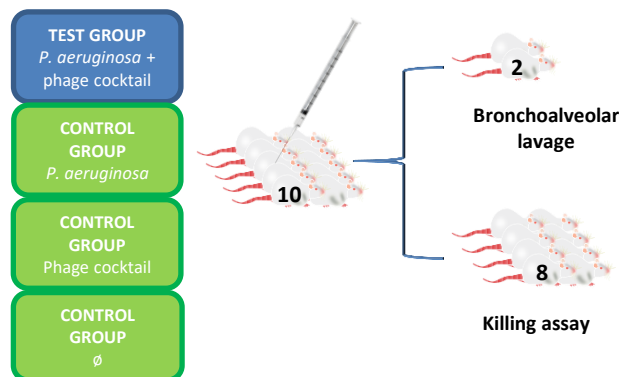


Figure4. Diagram of *in vivo* analysis and treatments.

EXPECTED RESULTS

- Reduction in viable cell numbers and total biomass in both early and mature biofilms.
- Significant increase in mice survival in the infected and treated group when compared with non-treated group.
- Cytokine and lactate dehydrogenase levels should be reduced, as well as bacteriophage and viable cell counts, when compared with non-treated group.

REFERENCES

1. Bodey, G. P., Bolivar, R., Fainstein, V., & Jadeja, L. (1983). Infections caused by *Pseudomonas aeruginosa*. Review of Infectious Diseases, 5(2), 279-313.
2. American Thoracic Society & Infectious Diseases Society of America. (2005). Guidelines for the management of adults with hospital-acquired, ventilator-associated, and healthcare-associated pneumonia. American journal of respiratory and critical care medicine, 171(4), 388.
3. Gu, J., Liu, X., Li, Y., Han, W., Lei, L., Yang, Y., & Sun, C. (2012). A method for generation phage cocktail with great therapeutic potential. PLoS One, 7(3), e31698.
4. Marinelli, L. J., Pluri, M., Swigořová, Z., Balachandran, A., Oldfield, L. M., van Kessel, J. C., & Hatfull, G. F. (2008). BRED: a simple and powerful tool for constructing mutant and recombinant bacteriophage genomes. PLoS One, 3(12), e3957.